



Hy-Laboratories Ltd.  
Park Tamar  
Rehovot 76326 Israel  
Tel : 972-8-9366475  
Fax : 972-8-9366474  
[hylabs@hylabs.co.il](mailto:hylabs@hylabs.co.il)  
[www.hylabs.co.il](http://www.hylabs.co.il)

# DTM AGAR

## Instructions for use

Cat No. PD109

DD019

DD027

BP272

DS034

### A) Preparation and collection of specimen.

1. Skin - Clean the affected area with 70% ethyl or isopropyl alcohol prior to removing skin scales. Remove scales from dry and peeling lesions by scraping from the inflamed edges towards the healthy skin with a sterile scalpel. Avoid using large scales as they should not be used as specimens. With acute inflamed or vesicular lesions the skin of the blister must be carefully removed with scissors and forceps. It should be cultured together with the contents of the blister, and if possible, scales from the surrounding areas. With infiltrates on granulomatous processes, collect material from the depth and from skin folds with a sharp spoon. Collect the material on a piece of filter paper or directly on the media.

2. Hair - Pluck the roots of dull, lusterless hair with forceps on the media.

3. Nails - Differentiate between two types of nail infections.

a) **Subungual Infection:** All grossly deformed surface parts of the nail are removed carefully with scissors, nail file or scalpel. Chips of nail are then collected from the nail bed.

b) **Surface Infection:** Nail chips or small dust like particles are scraped from the surface of the nail body. Do not inoculate the media with nail pieces cut from extracted nails. It is best to use a nail fraise.

4. Nail Wall: Collect drops of secretion produced by applying pressure to the nail wall.

### B) Inoculation

Distribute the material on the agar media with a sterile swab, loop or forceps. Slightly press the larger particles against the agar in order to obtain good contact between the specimen and the agar surface.

### C) TABLE OF RESULTS

Type of microorganism and incubation conditions of culture	MEDIA	
	Development	Inhibition
Bacteria ( 25-30°C up to 4 days)	-	+
Yeasts (25-30°C up to 4 days)	-	+
<i>S. cerevisiae</i> ATCC 2338	-	+
<i>C. albicans</i> ATCC 10231	+(no change background)	
<i>C. tropicalis</i>	-	+
Molds (25-30°C up to 4 days)		
<i>Aspergillus niger</i> ATCC 6275	-	+
<i>P. expansum</i> ATCC 7861	-	+
<i>P. notatum</i> ATCC 10108	±( no change of background)	±
Dermatophytes* 25°C		
<i>Trychophyton rubrum</i> (var KW)	+background becomes red	-
<i>Trychophyton floccosum</i>	+background becomes red	-
<i>Microsporum canis</i>	+background becomes red	-
<i>Microsporum gypseum</i>	+background becomes red	-
<i>Trychophyton rubrum</i> (var flava)	+background becomes red	-

\*The growth of dermatophytes leads to a color change of the medium underneath the colonies from rust to red bordeaux, even before the colonies are fully developed. As growth progresses the color of the whole medium will gradually become red and a white mycelium will become visible. Certain saprophytic mold contaminants may also grow on rust side, but the colonies and the mycelium appear before the medium's color change takes place. Certain cycloheximide-resistant yeasts may produce a color change of the medium, but they form smooth colonies without any mycelium.